

FORM PTO-1390
(REV. 10-95)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY DOCKET NUMBER
A-67653/DCA/DAV**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

U.S. APPLICATION NO. (if known, see 37 C.F.R. 1.5)

09/230955INTERNATIONAL APPLICATION NO.
PCT/GB97/02108INTERNATIONAL FILING DATE
5 August 1997PRIORITY DATE CLAIMED
5 August 1996

TITLE OF INVENTION

ASSESSMENT OF CERVICAL CELLS

APPLICANT(S) FOR DO/EO/US

MASON, Robert James; PASCOE, Edward William; HOLMES, Christopher Harold

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371
3. This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. has been transmitted by the International Bureau. (**see enclosed Form PCT/IB/308**)
 - c. is not required, as the application was filed in the United States Receiving Office (RO/US)
6. A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. have been transmitted by the International Bureau.
 - c. have not been made; however, the time limit for making such amendments has **NOT** expired.
 - d. have not been made and will not be made.
8. A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern other document(s) or information included:

11. An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. A **FIRST** preliminary amendment.
 - A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. A substitute specification.
15. A change of power of attorney and/or address letter.
16. Other items or information.

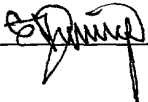
IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

300 Rec'd PCT 04 FEB 1999

In re application of:)	Examiner: Unknown
)	
Robert J. Mason, et al.)	Group Art Unit: Unknown
)	
National Phase Entry of)	
International Application No.:)	
PCT/GB97/02108)	
)	
International Filing Date:)	
August 5, 1997)	
)	
Serial No.: Unknown)	
)	
Filed: February 4, 1999)	
)	
For: ASSESSMENT OF)	
CERVICAL CELLS)	
_____)	

"EXPRESS MAIL" MAILING LABEL NUMBER EL 270 908 209 US
DATE OF DEPOSIT February 4, 1999
I HEREBY CERTIFY THAT THIS PAPER OR FEE IS BEING DEPOSITED
WITH THE UNITED STATES POSTAL SERVICE "EXPRESS MAIL POST
OFFICE TO ADDRESSEE" SERVICE UNDER 37 CFR 1.10 ON THE DATE
INDICATED ABOVE AND IS ADDRESSED TO:
ASSISTANT COMMISSIONER FOR PATENTS, WASHINGTON, DC 20231.

TYPED NAME Geody Domingo

SIGNED  _____

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents
BOX PCT
Washington, DC 20231

Sir:

Prior to substantive examination of the present case, Applicant offers the following amendments and remarks.

The Commissioner is authorized to charge any fees, including extension fees, which may be required, or credit any overpayment to Deposit Account No. 06-1300 (Our Order No. A-67653/DCA/DAV).

64050 56050

National Phase Entry of
International Application No.:
PCT/GB97/02108 which was filed:
August 5, 1997

In the claims:

Please amend the claims as follows:

4. (Amended) A method according to claim 1[or claim 2] wherein the specific binding substances include one or more such substances which complete for binding to cervical tissue with one or more antibodies obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 OJG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
9. (Amended) Use of a specific binding substance according to claim 6[or claim 7] for assessment of the nature or condition of cells of the cervix in a tissue sample.

Remarks

Applicant respectfully requests entry of these amendments. The Examiner is invited to contact the undersigned at (415) 781-1989 if any matter may be resolved by that means.

Respectfully submitted,

FLEHR HOHBACH TEST
ALBRITTON & HERBERT LLP



Dolly A. Vance
Reg. No. 39,054

Four Embarcadero Center
Suite 3400
San Francisco, CA 94111-4187
Telephone: (415) 781-1989

National Phase Entry of
International Application No.:
PCT/GB97/02108 which was filed:
August 5, 1997

APPENDIX OF CLAIMS:

1. A method of determining abnormality in a tissue sample containing cells of the cervix, the method comprising determining binding of specific binding substances to the sample and comparing the binding with the pattern of binding of said specific binding substances to a normal cervical cell sample.
2. A method according to claim 1 wherein the specific binding substances include one or more polypeptides each comprising an immunoglobulin antigen binding domain.
3. A method according to claim 2 wherein the specific binding substances include one or more polypeptides each comprising the immunoglobulin antigen binding domain obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 OJG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
4. (Amended) A method according to claim 1 [or claim 2] wherein the specific binding substances include one or more such substances which complete for binding to cervical tissue with one or more antibodies obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 OJG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
5. A hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 OJG, United Kingdom on 6 February 1995 ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
6. Use of a hybridoma of claim 4 and/or immunoglobulin antigen binding domain obtainable therefrom in the obtention of one or more specific binding substances of use in assessment of the state or condition of cervical cells in a tissue sample.
7. A specific binding substance including an immunoglobulin antigen binding domain obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 OJG, United Kingdom on 6 February 6, 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
8. A specific binding substance which competes for binding to cervical tissue with a specific binding substance according to claim 6.
9. (Amended) Use of a specific binding substance according to claim 6 [or claim 7] for assessment of the nature or condition of cells of the cervix in a tissue sample.

Applicant or Patentee: _____ Attorney's
 Serial or Patent No.: _____ Docket No.: _____
 Filed or Issued: _____
 For: "Assessment of Cervical Cells"

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY
 STATUS (37 CFR 1.9 (f) and 1.27 (c)) — SMALL BUSINESS CONCERN

I hereby declare that I am
 the owner of the small business concern identified below:
 an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN Quantum Biosystems Ltd (in Creditors Voluntary Liquidation)
 ADDRESS OF CONCERN acting by its liquidator, Roger William Gillett of Crane & Partners, Rutland House, 44 Masons Hill, Bromley, Kent, BR2 9EQ, U.K.

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9 (d), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention, entitled "Assessment of Cervical Cells" by inventor(s) Robert James MASON, Edward William PASCOE, Christopher Harold HOLMES described in

the specification filed herewith
 application serial no. _____, filed _____
 patent no. _____, issued _____

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9 (d) or by any concern which would not qualify as a small business concern under 37 CFR 1.9 (d) or a nonprofit organization under 37 CFR 1.9 (e).

*NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME _____
 ADDRESS _____
 INDIVIDUAL SMALL BUSINESS CONCERN NONPROFIT ORGANIZATION

NAME _____
 ADDRESS _____
 INDIVIDUAL SMALL BUSINESS CONCERN NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28 (b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING ROGER WILLIAM GILLETT (LIQUIDATOR)
 TITLE OF PERSON OTHER THAN OWNER FOR AND ON BEHALF OF
 ADDRESS OF PERSON SIGNING QUANTUM BIOSYSTEMS LIMITED
(IN LIQUIDATION)

SIGNATURE _____ DATE 25-01-1999

Messrs. CRANE & PARTNERS
CHARTERED ACCOUNTANTS
44 MASON'S HILL
BROMLEY, KENT. BR2 9EQ

ASSESSMENT OF CERVICAL CELLS

The present invention relates to assessment of cells in a sample of tissue containing cells of the cervix. More particularly, it relates to evaluation of the state of cells of the cervix, discriminating between normality and some deviation from normality, and is generally for use in screening women to detect those whose cervical cells are abnormal. Samples found to be abnormal may be examined in more detail and the condition of cells in the cervix investigated further. Identification of a malignant or pre-malignant condition is typically followed by appropriate treatment following more extensive diagnostic procedures.

Cancer of the cervix is the second most common cancer in women. The current method of detection is the Papanicolau or PAP test, which uses conventional cytological dyes to stain cells in a smear sample, enabling the visual detection of cellular nuclei and cytoplasm with a microscope. Trained personnel make semi-subjective assessments of the normality or otherwise of the cells examined. The PAP test, though universally accepted, is labour-intensive and prone to human error, as evidenced by a number of recent well-publicised scares casting doubt on the accuracy of assessments made by a few of those people who spend their entire day looking at smear samples.

An alternative or more objective way of

assessing the state of cells in cervical cell samples would be useful and advantageous. Benefit would be obtained by the removal or at least amelioration of problems arising from the need for subjective, visual
5 evaluation.

It is known that cells within tissues of the mammalian body express cellular markers that are either unique or partially restricted to particular cell populations. Different cell populations may be
10 distinguished by virtue of their individual cell markers. Thus, a cell may be shown to belong to a particular cell population (eg lymphoid cells) because of its expression of markers defining that population.

These cell markers may be various kinds of
15 molecules, including proteins, lipids, carbohydrates and combinations of these, such as glycoproteins, glycolipids and lipoproteins.

It is possible to detect cell markers using binding molecules, such as antibodies, with the
20 requisite specificity. Such binding molecules can be used in qualitative or quantitative detection of cells which bear particular markers and are thus included in a particular cell population.

Holmes et al have described previously a
25 monoclonal antibody able to bind specifically to an antigen on normal hepatocytes within the liver but not able to bind cells other than hepatocytes. No binding could be detected on a number of transplanted and

primary dimethylaminoazobenzene-induced hepatomas nor on liver cells from patients with a variety of liver diseases. (See: *Tumour Progression Markers - Proceedings of the Sixth Meeting of the European Association for Cancer Research, Budapest 12-15 October 1981*, 471-481 (1982); *Liver* (1983), 3: 295-302; *Int. J. Cancer* (1982), 29: 559-565; *Cancer Research* (1984), 44: 1611-1624.)

The present invention is founded in the realisation that it is possible to recognise a pattern of surface antigens on cells of the cervix which represents normality, such that a deviation from a determined and noted pattern of normality can be perceived. Conveniently, antibodies or other specific binding molecules may be used in the qualitative and/or quantitative detection of marker antigens on the cells, enabling increased or reduced expression or loss of one or more of the markers to be correlated with a disease (or pre-disease) state. In screening, this enables samples with some deviation from normality to be identified and examined further, ie suspect samples are highlighted for further examination by suitably qualified personnel. If abnormality in a particular sample is serious or potentially serious, appropriate steps may be taken to examine and then perhaps treat the woman from whom the sample was obtained. Diagnosis and decisions on the need for and nature of treatment remain the domain of

clinicians.

For operation of the present invention it is not necessary for the marker antigens actually employed in the test to be identified. Ultimately, what is
5 important is that a panel of antibodies or other binding molecules is identified as able to bind cervical cells with a pattern which is associated with the cervix being normal, and that deviation of binding of those binding molecules from the pattern of
10 normality is correlated with the onset of pathological conditions. The onset of pathology may then be identified in samples containing cervical cells by some deviation from the established pattern of normality for those particular binding molecules.

15 This is exemplified by data included herein relating to five monoclonal antibodies able to bind to various cells of the cervix. The pattern of binding of these antibodies to an abnormal tissue sample containing cervical cells is different from the
20 binding to normal cervical samples enabling identification of abnormality. Clearly, other specific binding molecules may be employed successfully in aspects of the present invention as long as the requirements set out in the preceding
25 paragraph are satisfied. Such other molecules may bind the antigens bound by the exemplified antibodies, at the same or different epitopes. Indeed they may bind different antigens altogether.

According to a first aspect of the present invention there is provided a method of determining abnormality in a tissue sample containing cells of the cervix, the method comprising determining binding of antibodies to the sample and comparing the binding with the pattern of binding of said antibodies to a normal cervical cell sample. The pattern of binding of the antibodies to normal cervical cells may, and generally will, be established prior to performance of the method.

The reactivities of antibodies on normal and test samples may be determined by any appropriate means. Tagging with individual reporter molecules is one possibility. The reporter molecules may directly or indirectly generate detectable, and preferably measurable, signals. The linkage of reporter molecules may be directly or indirectly, covalently, eg via a peptide bond or non-covalently. Linkage via a peptide bond may be as a result of recombinant expression of a gene fusion encoding antibody and reporter molecule.

One favoured mode is by covalent linkage of each antibody with an individual fluorochrome, phosphor or laser dye with spectrally isolated absorption or emission characteristics. Suitable fluorochromes include fluorescein, rhodamine, phycoerythrin and Texas Red. Suitable chromogenic dyes include diaminobenzidine.

Other reporters include macromolecular colloidal particles or particulate material such as latex beads that are coloured, magnetic or paramagnetic, and biologically or chemically active agents that can directly or indirectly cause detectable signals to be visually observed, electronically detected or otherwise recorded. These molecules may be enzymes which catalyse reactions that develop or change colours or cause changes in electrical properties, for example. They may be molecularly excitable, such that electronic transitions between energy states result in characteristic spectral absorptions or emissions. They may include chemical entities used in conjunction with biosensors. Biotin/avidin or biotin/streptavidin and alkaline phosphatase detection systems may be employed.

The mode of determining binding is not a feature of the present invention and those skilled in the art are able to choose a suitable mode according to their preference and general knowledge.

The signals generated by individual antibody-reporter conjugates may be used to derive quantifiable absolute or relative data of the relevant antibody binding in normal and test samples. In addition, a general nuclear stain such as propidium iodide may be used to enumerate the total cell population in a sampled smear, allowing the provision of quantitative ratios of individual cell populations relative to the

total cells in a smear, at least where individual antibody reactivities correlate with particular cervical cell populations.

An actual expansion or reduction in the absolute numbers of a particular cell population is not a necessary pre-requisite for the purposes of this invention. Any change in the detection of antibody binding (and by implication change in the phenotypic cell markers) that is detectable, and preferably quantifiable, relative to established parameters of normality, is of relevance.

In addition, non-epithelial cells such as leukocytes are known to infiltrate cervical tissue as a consequence of pathological conditions. These may be enumerated by use of readily available monoclonal antibodies against pan-leukocytic markers, thus providing a further level of analysis.

Antibodies which are specific for a target of interest may be obtained using techniques which are standard in the art. Methods of producing antibodies include immunising a mammal (eg mouse, rat, rabbit, horse, goat, sheep or monkey) with the protein or a fragment thereof or a cell or virus which expresses the protein or fragment. Immunisation with DNA encoding the target polypeptide is also possible. Antibodies may be obtained from immunised animals using any of a variety of techniques known in the art, and screened, preferably using binding of antibody to

antigen of interest. For instance, Western blotting techniques or immunoprecipitation may be used (Armitage et al, 1992, Nature 357: 80-82).

The production of monoclonal antibodies is well established in the art. Monoclonal antibodies can be subjected to the techniques of recombinant DNA technology to produce other antibodies or chimeric molecules which retain the specificity of the original antibody. Such techniques may involve introducing DNA encoding the immunoglobulin variable region, or the complementarity determining regions (CDRs), of an antibody to the constant regions, or constant regions plus framework regions, of a different immunoglobulin. See, for instance, EP184187A, GB 2188638A or EP-A-0239400. A hybridoma producing a monoclonal antibody may be subject to genetic mutation or other changes, which may or may not alter the binding specificity of antibodies produced.

As an alternative or supplement to immunising a mammal with a peptide, an antibody specific for a target may be obtained from a recombinantly produced library of expressed immunoglobulin variable domains, eg using lambda bacteriophage or filamentous bacteriophage which display functional immunoglobulin binding domains on their surfaces; for instance see WO92/01047. The library may be naive, that is constructed from sequences obtained from an organism which has not been immunised with the target or may be

one constructed using sequences obtained from an organism which has been exposed to the antigen of interest (or a fragment thereof).

Antibodies may be modified in a number of ways.

5 Indeed the term "antibody" should be construed as covering any specific binding substance having a binding domain with the required specificity. Thus this covers antibody fragments, derivatives, functional equivalents and homologues of antibodies,
10 including any polypeptide comprising an immunoglobulin binding domain, whether natural or synthetic. Chimaeric molecules comprising an immunoglobulin binding domain, or equivalent, fused to another polypeptide are therefore included. Cloning and
15 expression of chimaeric antibodies are described in EP-A-0120694 and EP-A-0125023.

It has been shown that the function of binding antigens can be performed by fragments of a whole antibody. Example binding fragments are (i) the Fab
20 fragment consisting of VL, VH, CL and CH1 domains; (ii) the Fd fragment consisting of the VH and CH1 domains; (iii) the Fv fragment consisting of the VL and VH domains of a single antibody; (iv) the dAb fragment (Ward, E.S. et al., Nature 341, 544-546
25 (1989)) which consists of a VH domain; (v) isolated CDR regions; (vi) F(ab')₂ fragments, a bivalent fragment comprising two linked Fab fragments (vii) single chain Fv molecules (scFv), wherein a VH domain

and a VL domain are linked by a peptide linker which allows the two domains to associate to form an antigen binding site (Bird et al, *Science*, 242, 423-426, 1988; Huston et al, *PNAS USA*, 85, 5879-5883, 1988); (viii) 5 bispecific single chain Fv dimers (PCT/US92/09965) and (ix) "diabodies", multivalent or multispecific fragments constructed by gene fusion (WO94/13804; P. Holliger et al Proc. Natl. Acad. Sci. USA 90 6444-6448, 1993).

10 Hybridomas able to produce monoclonal antibodies of use in the present invention have been deposited and represent individual aspects of the present invention, as do the monoclonal antibodies themselves. Thus, the present invention provides individually each 15 of the hybridomas deposited as ECACC 95020718, 95020716, 95020720, 95020717 and 95020719, and mutants, derivatives and descendants of each of these hybridomas, whether or not able to produce antibody of the same or altered specificity.

20 The present invention also encompasses use of the hybridomas and antibodies obtainable therefrom in the obtention of other antibodies of use in the assessment of the state/condition of cervical cells in a tissue sample, ie able to bind to an antigen found 25 on the surface of one or more cell types of the cervix. Such use may involve isolation of the antigen bound by any of the antibodies obtainable from the deposited hybridomas and use of the antigen in raising

further antibodies, eg by immunisation and/or screening of 'phage libraries, as discussed above. The antigen may, for example, be isolated by immunoprecipitation from cervical cell extracts and then
5 used as an immunogen eg for generation of further monoclonal antibodies, or in screening a 'phage library, as appropriate.

Additionally, nucleic acid encoding one of the antibodies may be isolated from any of the hybridomas
10 and used in a recombinant expression system to produce whole antibody, an antibody fragment or chimaeras of antibody/antibody fragment fused to another polypeptide (eg label such as a peptide tag or enzyme). As mentioned above, cloning and expression
15 of chimaeric antibodies are described in EP-A-0120694 and EP-A-0125023.

Recombinant expression of polypeptides, including antibodies and antibody fragments, is well-known in the art.

20 Systems for cloning and expression of a polypeptide in a variety of different host cells are well known. Suitable host cells include bacteria, mammalian cells, yeast and baculovirus systems. Mammalian cell lines available in the art for
25 expression of a heterologous polypeptide include Chinese hamster ovary cells, HeLa cells, baby hamster kidney cells and many others. A common, preferred bacterial host is *E. coli*.

Suitable vectors can be chosen or constructed, containing appropriate regulatory sequences, including promoter sequences, terminator fragments, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. For further details see, for example, *Molecular Cloning: a Laboratory Manual*: 2nd edition, Sambrook et al, 1989, Cold Spring Harbor Laboratory Press. Transformation procedures depend on the host used, but are well known.

Thus, the present invention extends to any antibody or antibody fragment able to bind an antigen to which any (ie one or more) of the antibodies obtainable from the deposited hybridomas is able to bind. Ability to bind the same antigen may be assessed, for example in a binding inhibition assay or in a band-shift assay on an electrophoretic gel.

The present invention also provides in a further aspect the use of any antibody or antibody fragment of the invention, including those obtainable from any of the deposited hybridomas, in assessment of the nature or condition of cells of the cervix in a tissue sample, as disclosed.

Further aspects and embodiments of the present invention will be apparent to those skilled in the art.

Before illustrating embodiments of the present invention in more detail by way of example, it is

helpful to understand the organisation of the cervix and the relationships between the different cell-types found there.

5 *Cell populations of the cervix in normal and pathological conditions*

The cervix is essentially composed of two distinct cell types: the squamous epithelium and the columnar epithelium, each of which is located in an anatomically distinct region of the tissue. The squamous epithelium is located at the exterior aspect (the exocervix) of the cervical opening (the cervical os), while the columnar epithelium extends into the endocervical canal (the endocervix). These two distinct epithelial cell types come into contact in the vicinity of the cervical os, at the squamo-columnar junction. The squamo-columnar junction is of clinical importance as it is the region where the majority of malignancies arise. For diagnostic validity, a cervical smear sample must include cells from this region. In order to ensure that this has been achieved, a smear must contain columnar as well as squamous epithelial cells.

Columnar cells are the source of cervical mucous. They are generally arranged as a single cell layer lining the endocervix, which is thrown into deep folds to form the cervical glands. A small proportion (5%) of cervical tumours are derived from columnar

cells i.e. the adenocarcinomas.

In some cervixes, a cuboidal cell layer, the so-called reserve cell population, is disposed beneath the columnar cells. The role of reserve cells is unclear, but believed by many investigators to give rise to columnar epithelium.

In contrast to the columnar epithelium, the squamous epithelium, from which most (95%) cervical tumours arise, is a multilayered dynamic stem cell system under constant renewal.

The stem cell compartment itself is located adjacent to the basement membrane within the basal cell layer. Stem cell division gives rise to parabasal, intermediate, and superficial cell derivatives. These are conventionally defined in terms of both their characteristic morphology and location within the squamous epithelium. The transition from basal cells located in the deepest layer of the squamous epithelium, to superficial cells at its surface is associated with progressive differentiation and a loss of proliferation until superficial squamous epithelial cells at the cervical surface are terminally differentiated.

The transition zone (TZ), adjacent to the squamo-columnar-junction is of clinical importance, as it includes a region of metaplastic squamous-epithelium. This is generated at puberty, in response to the acidic environment of the vagina. The data

presented herein quantify the proportion of samples where the TZ is present.

EXEMPLIFICATION OF EMBODIMENTS OF THE PRESENT

5 *INVENTION BY WAY OF ILLUSTRATION AND NOT LIMITATION*

Abbreviations:

ATCC - American Type Culture Collection; CIN - Cervical intra-epithelial neoplasia; CHAPS (3- [Cholamidopropyl)-dimethylammonio]-1-propanesulfonate);
10 CD - Cluster Designation nomenclature of established cell markers; DAB - Di-amino benzidine; ECACC - European Collection of Animal Cell Cultures; EDTA - Ethylene di-amine tetra acetic acid; HLA - Human
15 Leukocyte Antigen; HRPO - Horse radish peroxidase; Ig - Immunoglobulin; kDA - Kilo dalton; Mab(s) - Monoclonal antibody(ies); Mwt - Molecular weight; NS1 - Non-secretor 1; PAGE - Polyacrylamide gel electrophoresis; PBS - Phosphate buffered saline; PAP
20 - Papanicolau test; SDS - Sodium dodecyl sulphate; TBS - Tris buffered saline; TZ - Transition zone; W/V - Weight/volume ratio.

Cell markers

25 The work disclosed herein shows that under normal conditions, cell surface markers may be gained or lost as cells within a given lineage proceed along their differentiation pathway. Therefore, monoclonal

antibodies or other binding molecules exhibiting specific reactivities against these markers provide a means of monitoring the normal progression of cells along their differentiation pathway. In pathological conditions, the normal expansion or loss of these markers may be perturbed. Consequently, an expansion or loss of a cell surface marker is detectable as a particular cell population proliferates or becomes arrested at a discrete stage in its differentiation pathway.

The antibody reactivities described herein illustrate the situation where an overlapping continuum of reactivities is observed throughout normal squamous epithelial cell differentiation in the human cervix.

The designated nomenclature of the five monoclonal antibodies exemplified herein is:

1. CV3.6B5 / F3 / C2, hybridoma deposited as ECACC
95020718
2. 2C7 / B4 / D6., hybridoma deposited as ECACC
95020716
3. CV5.9G5.C6, hybridoma deposited as ECACC
95020720
4. HG3 / E11 / C4, hybridoma deposited as ECACC
95020717
5. BC4 / E7 / E5, hybridoma deposited as ECACC
95020719

Generally herein, these antibodies are referred to in their abbreviated forms. The hybridomas were deposited at European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 0JG, United Kingdom on 6 February 1995.

Thus:

- Basal cells are 6B5+ BC4- 9G5- HG3- 2C7-
- 10 Parabasal cells are 6B5+ BC4+ 9G5-HG3-2C7-
- Intermediate cells are 6B5- BC4+ 9G5+HG3+2C7-
- Superficial squames are 6B5- BC4- 9G5+HG3+2C7-
- Columnar cells are 6B5+ BC4- 9G5-HG3- 2C7+.

15 These characteristic antibody reactivity profiles reproducibly define discrete stages in the differentiation of squamous epithelial cells. Additionally, they readily allow columnar epithelial cells to be distinguished from squamous epithelial

20 cells.

Brief Description of Results

Immunoprecipitation of the 6B5 target protein from the cell surface of a cervical carcinoma cell line and from membrane preparations of cervical squamous epithelium:

- (a) Cell surface proteins on the cervical

carcinoma cell line C4II (Auersperg 1969. *J. Natl. Cancer Inst. USA* 43 151-173) were radioiodinated by the lactoperoxidase method (Marchalonis 1969. *Biochem J.* 113 299-305). Cells were solubilised with the
5 detergent CHAPS (3-[3-Cholamidopropyl]-dimethylammonio]-1-propanesulfonate).

Immunoprecipitations were carried out with test Mab against equal volumes of lysate (Houlihan et. al. 1992 *J. Immunol.* 149 668-675). The results show that Mab
10 6B5 detects a cell surface dimeric protein with components of approx. 181 and 184 kDa in a cervical epithelial cell line.

(b) Standard membrane extracts were prepared from sheets of cervical squamous epithelium (Evans
15 1979. *Laboratory techniques in biochemistry and molecular biology, Eds. Work and Work* 7 1-266, Elsevier). Membranes were radioiodinated using Iodobeads (Markwell 1982. *Anlyt. Biochem.* 125 427-432) and solubilised in CHAPS. Immunoprecipitations
20 were carried out as described above. The results show that a similar form of this protein is also present in a membrane-associated fraction of cervical squamous epithelium shown in (a).

Immunoprecipitated material was analysed by SDS-
25 PAGE under reducing conditions and autoradiography (Laemmli 1970. *Nature* 227 133-681).

Affinity isolation of the 6B5 target from C4II cells:

Affinity chromatography was performed as described by Houlihan et al. 1992 (*J. Immunol.* 149 668-675). Purified 6B5 antibody was conjugated to protein G-sepharose and cross-linked with dimethyl pimelimidate. 2 x 10⁸ C4II cells were solubilised in the detergent CHAPS. The lysate was passed through a preclearing column containing mouse Ig-sepharose and protein A sepharose (Sigma) and then through the 6B5 antibody column. Both columns were washed with 10 bed volumes of lysis buffer and then eluted with 50 mM triethylamine, pH 11.5. The eluate was neutralised with 2 M glycine pH 2.0, concentrated by microultrafiltration and analysed by SDS-PAGE under reducing conditions and coomassie blue staining.

A single 180 kDa product was specifically isolated by the 6B5 column and detected in reduced eluates. This component was not observed in the eluate representing the negative control - mouse Ig/protein A sepharose column. Immunoglobulin H-chain which had leached during elution from the negative control column was present, however, in the mouse Ig/protein A sepharose eluate.

Immunoblotting of detergent-solubilised endocervical material using antibody 2C7:

Endocervical tissue was minced in Hanks buffered saline and proteins were solubilised in CHAPS (3-[3-Cholamidopropyl]-dimethylammonio]-1-propanesulfonate)

buffer. The detergent soluble material was concentrated by microultrafiltration using a 30 kD cut-off filter. Proteins were electrophoretically separated on 1% agarose gels in Laemmli sample buffer
5 under reducing conditions. Proteins were transferred to microporous membranes for immunoblotting. Identical strips were probed with Mab 2C7 and tissue culture supernatant as a negative control. Parallel strips were stained with periodic acid-Schiff's reagent
10 which stains glycoproteins, and coomassie blue as a general protein stain (*Methods were based on those of Morales et. al. 1993 Human Reproduction 8 78-83*). -ve control immunoblot employed tissue culture supernatant.

15 Mab 2C7 specifically detects components migrating in the high m.wt. range that correspond to material stained by periodic acid-Schiff's reagent. The large m.wt (>500 kDa) of this material and detection by periodic acid-Schiff's reagent is
20 consistent with the presence of mucins. The Mab showed no reactivity with any proteins in the significantly lower m.wt. material stained by coomassie blue.

25 Immunoblotting of detergent-solubilised cervical epithelium using Mab 9G5:

Squamous epithelial sheets were isolated from normal cervixes using the enzyme Dispase (Boehringer).

Sheets were solubilised in the detergent CHAPS (3-[3-Cholamidopropyl)-dimethylammonio]-1-propanesulfonate). Cytokeratin-enriched extracts were prepared from the CHAPS-insoluble material by the method of Franke et al. 1981 (*Exp. Cell Res.* 131 209-213). Both
5 detergent (D) and cytokeratin (C) extracts were electrophoretically-separated by SDS-PAGE under reducing conditions (Laemmli 1970. *Nature* 227 133-681) and proteins were transferred to microporous
10 membranes for immunoblotting (Towbin et al. 1979. *Proc. Natl. Acad Sci USA* 76 4350-4354).

Mab 9G5 detects a single 40 kDa product in detergent-solubilised extracts but not in cytokeratin extracts of cervical squamous epithelium. A pan-
15 reactive cytokeratin Mab AH3 (*J.M. Houlihan Ph.D. thesis University of Bristol 1993*) was used as a control; keratins are detected in the cytokeratin extract.

20 Affinity chromatographic isolation of the 9G5 target protein from human amnion cells:

Affinity chromatography was performed as described by Houlihan et al. 1992 (*J. Immunol.* 149 668-675). Purified Mab 9G5 was conjugated to protein
25 G-sepharose and cross-linked with dimethyl pimelimidate. 2×10^8 amnion cells were isolated from the term placental membranes by incubation in trypsin followed by collagenase/hyaluronidase by the method of

Holmes et al. 1990. (*J. Immunol.* 144 3099-3015) and solubilised in the detergent TX-100. The lysate was passed through preclearing columns, comprising mouse Ig and protein A sepharose and then the Mab 9G5
5 column. The Mab 9G5 column was eluted with 50 mM triethylamine, pH 11.5. The eluate was concentrated by microultrafiltration and analysed by SDS-PAGE under non-reducing conditions and coomassie blue staining.

A single 40 kDa product was isolated from amnion
10 cell lysates by the Mab 9G5 affinity column. A fraction of this eluate was examined by immunoblotting with the 9G5 Mab and an irrelevant IgG1 Mab as a negative control. The 9G5 reacted with the 40 kDa product. The strip probed with the negative control
15 Mab was blank.

Immunoblotting of detergent-solubilised cervical epithelium using Mab HG3:

Squamous epithelial sheets were isolated from
20 normal cervixes using the enzyme Dispase (Boehringer). Sheets were solubilised in the detergent CHAPS (3-[3-Cholamidopropyl)-dimethylammonio]-1-propanesulfonate). Equal amounts of the CHAPS-soluble material were electrophoretically-separated by SDS-PAGE (*Laemmli*
25 1970. *Nature* 227 133-681) under both non-reducing and reducing conditions. Proteins were transferred to microporous membranes for immunoblotting (*Towbin et al.* 1979. *Proc. Natl. Acad. Sci. USA* 76 4350-4354).

Parallel strips tested with Mab HG3; tissue culture supernatant was used as a negative control.

The negative control blot was blank.

(NR - non-reduced; R - Reduced)

5 Mab HG3 detects a product of approx. 180 kDa, under both non-reducing and reducing conditions, in detergent soluble cervical squamous epithelium.

10 Immunoblotting of detergent-soluble and detergent-insoluble cervical epithelium using Mab BC4:

Squamous epithelial sheets were isolated from normal cervixes using the enzyme Dispase (Boehringer). Sheets were solubilised in the detergent CHAPS (3-[3-Cholamidopropyl)-dimethylammonio]-1-propanesulfonate).
15 CHAPS-soluble and CHAPS-insoluble material was adjusted to the same volume. Equal amounts of both soluble and insoluble fractions were electrophoretically-separated by SDS-PAGE (*Laemmli 1970. Nature 227 133-681*). The detergent soluble
20 fraction was analysed under both non-reducing and reducing conditions while the detergent insoluble fraction was examined under reducing conditions. Proteins were transferred to microporous membranes for immunoblotting (*Towbin et al. 1979. Proc. Natl. Acad Sci USA 76 4350-4354*).
25 Parallel strips were tested with Mab BC4; tissue culture supernatant was used as a negative control.

The negative control blot was blank.

Mab BC4 detects a protein of 200-210 kDa in non-reduced, but not in reduced, detergent soluble extracts of cervical squamous epithelium. No product was detected in detergent insoluble fraction of
5 cervical squamous epithelium.

Cervical Biopsies

Most of the data herein has been obtained from extensive investigations undertaken on biopsy
10 material, for the following reasons:

(a) A single biopsy specimen provides several serial tissue sections (each 5 um thick) for microscopic examination. All sections are therefore almost identical, and the various cell populations located in
15 the same positions relative to each other. Similarity in anatomical structure, at the cellular level, is therefore maintained between samples.

This enables reactivities of different antibodies to be investigated and compared, on the
20 same cell populations, between samples.

Different cell populations within a tissue are identified by their individual morphologies and locations. Therefore, the reactivities of individual antibodies against such cells in a biopsy sample,
25 enables their specificities to be determined and established.

(b) Pathological changes are accompanied by a disruption of the normal histology of the tissue.

Therefore, antibody reactivities can only be effectively investigated (and correlated with pathological conditions) in samples, where the tissue architecture is representative of the situation *in vivo*.

Pathological changes that precede malignancy are classified according to an established system of grading: CIN I, II and III. The progression of disease leads to carcinoma *in situ* and finally a frank tumour.

(c) Most importantly, the use of biopsy material was necessary to determine whether modulation of expression of the antibody targets occurred as a function of disease in the cervix. The profiles of antibody reactivities on tissue sections of normal biopsies provided a framework to establish their corresponding reactivities on abnormal smears. Such reactivities on normal and abnormal biopsies enabled the selection of antibodies for use in a cervical smear screening system, in accordance with the present invention.

Cervical Smears

The cells in a smear sample do not maintain their topographical relationships to each other, as in a tissue section of a biopsy. Antibodies selected on the basis of their specificities against various cell populations (as above), were investigated for their

ability to provide qualitative or quantitative information on normal and pathological smear samples. Their absolute or relative numbers, enumerated with these antibodies, provides a means of determining
5 detectable changes in these cell populations.

Tissue Distribution

Although the monoclonal antibodies were raised against cervical epithelium, they would be expected to
10 react against their target epitopes if present on non-cervical epithelium of shared embryological origin. Therefore, their tissue distribution in other epithelial tissue was determined, enabling further characterisation at that level.

15

Biochemical Data

These have been mainly derived using extracts of cervical amnion and placental tissue, electrophoretically separated on SDS-PAGE gels under
20 reducing or non-reducing conditions. Western blots of the resolved components were then probed with the panel of monoclonal antibodies, enabling their individual target epitopes to be defined at the molecular level.

25

The detection of such resolved components under defined experimental conditions provides information on the molecular structure of the target antigen. If detected under both reducing and non-reducing

conditions, it can be deduced to be a non-
conformationally dependent monomeric structure. On
the other hand, if only detected under non-reducing
conditions, the epitope is present on a
5 conformationally dependent structure.

The methods of extraction enable the identified
material to be assigned to known classes of cell
markers. For example, use of detergents do not
provide extracts with appreciable amounts of
10 cytokeratins. The cytokeratins are a complex family
of cytoplasmic filamentous protein structures that are
well documented in the scientific literature. They
are biochemically and antigenically related to varying
degrees, and are expressed in different epithelia in
15 different combinations of polypeptides. A given
epithelium or epithelial cell can therefore be
characterized by the specific pattern of its
cytokeratin components [Ref: Moll R et al, 1982].

Furthermore, with the exception of Mab 9G5, the
20 molecular weights of the target antigens precluded
them from belonging to the cytokeratin family. A
monoclonal antibody (AH3) with pan-Cytokeratin
reactivity was used to confirm that the 9G5 epitope
was a non-cytokeratin cell marker. This antibody has
25 been previously described [Ref: Houlihan 1993] .

Where sufficient or suitable biopsy material was
not obtainable, established carcinoma cell lines
derived from cervical tissue were utilised to prepare

such extracts.

Trypsin sensitivity.

The susceptibility of target epitopes to
5 proteolysis by trypsin was determined. The enzyme's
pronounced substrate specificity (restricted to lysine
and arginine residues) provides a means of
characterising the target epitopes further.

10 *Materials and Methods*

Standard buffers and reagents were prepared in
accordance with established procedures that are well
documented in the scientific literature so need not be
detailed here. Unless otherwise stated, all chemicals
15 were obtained from Sigma Chemical Co, Poole, Dorset,
UK.

Preparation Of Immunogen And Immunisation

Three sources of cervical cells: (A) smear
20 samples, (B) hysterectomies and (C) premalignant CIN
biopsies were evaluated for their potential use as
immunogen:

(A)

Cervical smears from routine samples were
25 dispensed into sterile PBS and washed twice. The
yield, composition and viability of cells in smears
taken from different sites within the cervix was
examined. The cellular composition of conventional

smears taken from the exocervix using a spatula, was compared with endocervical smears taken with a brush. Yields of squamous cells in exocervical smears varied; obtaining up to 10^6 , with viabilities of 50-60%.

- 5 Endocervical smears gave much lower yields of typically 10^4 , which also contained predominantly squamous cells.

Cytospin preparations of cervical smear samples were also examined immunohistologically and identified using anti-cytokeratin antibodies. The overwhelming majority were squames; the preparations contained few basal, parabasal, or columnar epithelial cells. It was concluded that cervical smears did not represent an effective source of material for use as immunogen, since they did not contain a sufficient cross section of cervical epithelial cell populations.

10
15

(B)

Total hysterectomy specimens are typically obtained from women with menorrhagia or fibroids; in these, the cervix is essentially normal. Such specimens are typically in the perimenopausal (38-45 yr old) period. The optimal method for obtaining single cell suspensions from whole normal cervical biopsy material was determined to be as follows.

20

25 The excess stromal tissue was removed with a scalpel. The fragments of tissue were floated in a solution of the enzyme Dispase II (1.2 units/ml, Boehringer Mannheim, Sussex, UK) in HANKS buffered

saline lacking $\text{Ca}^{2+}/\text{Mg}^{2+}$ overnight at 40C. This procedure disrupted the epithelial/stromal junction, such that sheets of epithelial cells could be gently teased apart. They were washed by low speed
5 centrifugation in HANKS buffered saline lacking $\text{Ca}^{2+}/\text{Mg}^{2+}$ and resuspended in 0.05% trypsin/0.02% EDTA (both w/v). After a 30 minute incubation at 37°C with stirring, trypsinization was arrested by the addition of 1.3 mg (in 5 ml of saline) of soya bean trypsin
10 inhibitor.

The secondary enzyme digestion with trypsin disaggregated the epithelial sheets to generate a single cell suspension.

Large fragments were allowed to settle out under
15 gravity and the supernatant was removed. The cell suspension was drawn through a 16 gauge needle to break up cell clumps, and filtered through a 100 um gauze. Cells were washed twice in PBS prior to use as immunogen.

20 (C)

In addition to normal cervical biopsies, immunogen from pathological CIN II/III biopsies were also used. The material was treated similarly, except for the initial overnight enzyme treatment also being
25 in trypsin.

The immunisation regime consisted of an initial intra-peritoneal inoculation of 5×10^5 cells as a priming dose, followed by five further immunisations

of $0.5-2 \times 10^6$ normal epithelial cells, at 2-3 weekly intervals in 8 week old female Balb/c mice.

As some proteolytic cleavage of cell surface components was possible, the effect of enzyme
5 treatment on the integrity of epithelial cell surface markers was ascertained. This was determined by monitoring its effect on CD44, CD55 and HLA Class 1 antigens, all of which are expressed on cervical epithelial cells. These established cell surface
10 markers can be detected by appropriate monoclonal antibodies which are readily commercially available [Ref: Knapp W, 1989]. These markers were found to be lost or reduced as a consequence of prolonged exposure to trypsin at 37°C. However, adopting measures to
15 minimise such effects i.e. reduced exposure at 37°C and use of the enzyme dispase in the initial overnight incubation at 4°C instead, ensured that the phenotypic profile of the disaggregated cells was maintained very closely to that of native cervical epithelium.

20

Fusion And Assay Of Supernatants From Hybridomas

Splenocytes from the immunised mice were fused with NS1 murine myeloma cells, and hybrids selected by conventional methodology that is described in a
25 number of publications [Ref: Kennet R et al, 1980, and Schrier M et al, 1980].

Candidate monoclonal antibodies were selected on the basis of their reactivities on tissue sections of

cervical biopsies using established immunohistological techniques [Ref: Holmes CH et al, 1990].

Briefly, sections were cut from frozen tissue blocks in a cryostat at 5 μ m thickness, thawed, air
5 dried at room temperature for 1 hour, fixed in ice-cold acetone for 10 mins and immunostained by an indirect immunoperoxidase technique. Supernatants, from wells containing growing hybridomas, were incubated for 45 mins at room temperature on tissue
10 sections. After washing in TBS for 5 minutes, they were incubated for 30 mins with a commercially available HRPO-conjugated rabbit anti-mouse Ig reagent, optimally diluted in TBS containing 10% normal human serum. After two more washes, sections
15 were developed with DAB and hydrogen peroxide for 5 mins. After stopping the reaction by washing the slides in tap water for 5 mins, the sections were counterstained in haematoxylin, dehydrated, cleared in HistoClear and mounted in DPX mountant.

20 Candidate hybridomas, secreting antibodies of interest, were cloned to stability by the method of limiting dilution. The antibodies secreted by such clones were reassayed to confirm antibody specificity by immunostaining, as above.

25 Ig production by hybridomas was also screened by an ELISA technique, using commercially available reagents from Dako AS, Copenhagen, Denmark): Rabbit anti-mouse Ig (product no: Z259) optimally diluted at

1/2000 was used as the solid phase capture reagent. Supernatants from hybridomas were incubated for 60 min; bound antibody was detected with a HRPO-conjugated rabbit anti-mouse Ig (product no: P260) diluted 1/1000 incubated for 45 mins. Both incubations were at room temperature; the reagents were diluted in PBS, and the wells washed between incubations with PBS-0.025% Tween.

10 *Biochemical Characterisation*

The methods utilised are generally well known and documented in published laboratory technical manuals [Ref: Harlow E and Lane D, 1988 and Work and Work, 1979] and in individual publications [Ref: Marchalonis, 1969; Markwell, 1982; and Laemmli, 1970]. Detailed descriptions of specific procedures undertaken in these investigations have been previously published [Ref: Holmes CH et al 1990 and Houlihan JM et al, 1992].

20

EXAMPLE 1 - Mab 6B5

This antibody of isotype IgG1 was raised against an immunogen prepared from squamous epithelial cells isolated from the cervixes of hysterectomy specimens.

25

Reactivity

Its specificity is mainly against parabasal and basal cells in cervical squamous epithelium together

with basement membrane. However, it also crossreacts with columnar epithelium and non-epithelial stromal elements.

Although parabasal cells are not present in large numbers in normal smears, the antibody does identify the small number that are present; based cells are usually not present in smear samples. However, 6B5 reactivity increases significantly in pathological conditions. With tumours, 6/7 squamous cell carcinomas showed antibody reactivity. This reactivity was retained on one adenocarcinoma; this was expected, as 6B5 also reacts with normal columnar cells. There is also an expansion of 6B5 reactivity in CINs: in 9/15 moderate to severe CIN specimens tested, there was a marked expansion in the affected squamous epithelium relative to the native unaffected epithelium.

The anti-stromal reactivity is only of relevance in biopsy material, as stromal elements are not present in smear samples. The anti-columnar reactivity is not expected to interfere in the intended mode of application, as adequate controls exist to correct for this effect (see reactivity of Mab 2C7).

25

(a) *Normal Cervical Epithelium*

No of tests: 90; No of patients: 44 (with TZ:

21)

Parabasal cells and basement membranes were intensely reactive in squamous epithelium. Columnar cells were also positive, with reactivity expanded in the transformation zone in 21/21 specimens.

5 (b) *Premalignant Cervical Epithelium (CIN II/III or III)*

No of tests: 42; No of patients: 30 (with CIN: 15)

The reactivity was expanded in 9/15 CIN
10 specimens.

(c) *Cervical Carcinomas*

The antibody reacted strongly with 5/7 squamous cell carcinomas and with both adenocarcinomas tested.

15 *Tissue Distribution*

Despite its restricted distribution in the cervix, the target epitope is present in other epithelial tissue:

Placenta:

20 Epithelial cells in term placental membranes (amnion and cytotrophoblasts) are positive. The basement membrane beneath the syncytiotrophoblast is positive. In first trimester placenta, both villous cytotrophoblast and syncytiotrophoblast are positive.
25 The antibody shows differential activity with cytotrophoblast in extravillous cell columns: cells at the base of these columns are positive while those at the periphery are negative.

Kidney: glomeruli +; tubules -
Pancreas: ducts and acini +; lamina propria -
Colon: epithelium +; lamina propria -
Liver: hepatocytes +; bile duct/mesenchymal
5 cells -
Endometrium: glandular epithelium -; lamina
propria -; myometrium -; arteries -
Epidermis: parabasal cells +; basement membrane +
(foreskin) basal cells + / -; intermediate cells -;
10 superficial cells -

Biochemical

Because of its limited reactivity in the normal
cervix, an established cervical carcinoma cell line
15 C4II was used to characterise this antibody
biochemically. The C4II cell line [Ref: Auersperg N
and Hauvryl AP, 1962 and Auersperg N, 1969] obtained
from ECACC, Porton Down, UK is well documented in the
scientific literature. It is also deposited in the
20 ATCC under Accession No: CRL 1595.

Mab 6B5 detects a dimeric product of
approximately 181-184 kDa, by immunoprecipitation on
the cell surface of C4II. A single component having
this molecular weight has subsequently been
25 immunoprecipitated directly from radiolabelled
membrane preparations of normal *in vivo*-derived
cervical squamous epithelial cells. However, the
antibody does not detect these components on cervical

extracts by western blotting. Taken together, these data indicate that Mab 6B5 detects a conformationally-dependent epitope on a cell surface (non-cytokeratin) protein. The 6B5 target has also been isolated to a high degree of purity by immunoaffinity chromatography, for N-terminal sequencing.

Trypsin sensitivity

The reactivity of the antibody is maintained with the C4II cell line after a short 10 min exposure to trypsin at 0.05% (w/v). However, antibody reactivity with placental membranes or amnion cells is abolished after a longer 1 hr exposure to trypsin at 0.1% (w/v). The antibody target is therefore only partially resistant to trypsin.

EXAMPLE 2 - Mab 2C7

This antibody of isotype IgG1 was raised against an immunogen prepared from normal cervixes of hysterectomy specimens.

Reactivity

The antibody reacts specifically and solely with columnar epithelial cells.

(a) Normal Cervical Epithelium

No of tests: 83; No of patients: 43 (with TZ:

24)

This antibody reacted specifically with columnar epithelial cells, and not with any other cell population in the cervix. The squamous epithelium was negative in all cases.

5 (b) *Premalignant Cervical Epithelium (CIN II/III or III)*

No of tests: 42; No of patients: 30 (with CIN: 15)

No reactivity with either normal or neoplastic epithelium; only adjacent columnar epithelium was stained.

(c) *Cervical carcinomas*

The antibody did not react with 7/7 squamous carcinomas tested, as expected. However, it reacted with one of the two adenocarcinomas tested.

It can be concluded that the target epitope of Mab 2C7 is solely restricted to columnar cells, and not expressed in either normal squamous epithelium or in CIN lesions. The antibody is considered useful since the presence of columnar cells on a smear indicates that the squamo-columnar junction has been sampled, and hence that the smear is patent (or adequate). A correctly sampled smear therefore contains between 1-5% of 2C7 reactive cells.

25 The 2C7 target epitope is distinct from that of 6B5. In normal smears, both antibodies identify the columnar cell population. However, whilst 6B5 additionally reacts with parabasal cells, 2C7 only

detects columnar cells. Therefore, together, they provide a means of enumerating both cell populations by analysis of their individual reactivity profiles.

5 *Tissue Distribution*

10	Placenta:	The amnion and cytotrophoblast in placental membranes, and syncytiotrophoblast were negative in first trimester placentae and at term.
	Pancreas:	Epithelium of ducts +
	Colon:	Glandular epithelium +
	Tonsil:	Stratified epithelium -
	Epidermis:	Foreskin -
15	Liver:	Hepatocytes- ; bile duct +/-
	Endometrium:	Epithelium -

Biochemical

20 Detergent extracts of endocervical epithelium were prepared by incubating fragments of endocervix in buffer containing CHAPS. On Western blots of material separated by SDS-PAGE, the antibody reacted with unresolved high Mwt components of >400,000 kDA in these extracts. This was further investigated by

25 resolving the extracted material on 1% agarose gels, which are more suitable than SDS-PAGE gels for larger molecules. The antibody reacted with components that precisely corresponded to a fraction that also stained

with periodic acid-Schiff's base, but not with Coomassie Blue. This indicated that the target epitope represented a mucin or a mucin-associated product.

5

Trypsin sensitivity:

The target epitope appears to be trypsin insensitive, as the antibody reacted with cell preparations of cervical tissue which had been trypsinised for periods of up to 1 hr.

EXAMPLE 3 - Mabs 9G5 and HG3

Both antibodies are of isotype IgG1.

Mab 9G5 was raised against an immunogen prepared from squamous epithelial cells isolated from normal cervixes of hysterectomy specimens.

Mab HG3 was raised against an immunogen prepared from cervical biopsies of pathological CIN material.

20 *Reactivity*

Both antibodies react primarily with superficial and intermediate squamous epithelium in the cervix.

(a) *Normal Cervical Epithelium*

25 No of tests: 83; No of patients: 44 (with TZ: 21 for Mab 9G5 and 24 for Mab HG3)

Both antibodies have similar patterns of reactivity in the normal cervix, with specificities

against superficial and intermediate squamous
epithelial cells. They are unreactive with basal
cells; however, Mab 9G5 reacts with parabasal cells in
some specimens, where the reactivity fades within the
5 upper parabasal layer. In contrast, Mab HG3 does not
react with parabasal cells in any of the specimens.

Mab 9G5 does not react with columnar cells;
however, Mab HG3 shows weak reactivity on columnar
cells in a small number of specimens.

10

(b) *Premalignant Cervical Epithelium (CIN II/III or
III)*

No of tests: 42; No of patients: 30 (with CIN:
15)

15

In general, both antibodies show detectable
differences between normal and abnormal cervical
epithelium. The reactivity is modulated in CINs,
where it is either reduced or absent; e.g. the
reactivity of Mab 9G5 is markedly reduced in 10/15
20 specimens. In these lesions, the depth of immuno-
staining (denoting antibody reactivity) is reduced in
terms of the number of cell layers, when compared to
normal squamous epithelium.

25 (c) *Cervical Carcinomas*

Both antibodies differed in their reactivities
against the seven squamous cell carcinomas tested:

Mab 9G5 was completely unreactive with 3/7

specimens tested. The remaining 4 tumours showed heterogenous reactivity.

Mab HG3 showed extensive reactivity with 3/7 of the squamous cell carcinomas, some reactivity with 2,
5 and was unreactive with the remaining 2.

Heterogeneity is defined as areas of both reactivity and unreactivity within a particular specimen. Generally, despite similar reactivities in the normal cervix, the reactivity of Mab 9G5 with
10 these tumours was more limited than that of Mab HG3. In particular, Mab 9G5 was negative with two tumours which Mab HG3 reacted extensively with.

When tested on two adenocarcinomas, Mab HG3 reacted with both tumours; whereas Mab 9G5 was
15 unreactive against both.

Mabs 9G5 and HG3 recognise distinct target epitopes on the same cell populations. However, it is considered desirable to utilise them in tandem, for the following reasons:

20 (a) It is not known whether all cells that comprise the intermediate and superficial cell population display both target epitopes. Even if they normally do, their expression may modulate with stages of the cell or oestrus cycles.

25 (b) As superficial epithelial cells are a mainly dead or dying cell population, it is conceivable that their surface markers would be heterogenous in their detectability.

(c) Cervical smears are largely comprised of superficial and intermediate cells; with columnar and parabasal cells being a minority. Consequently, a decisive factor in deriving conclusions regarding the clinical status of the sample, would be based on the information concerning these cells.

For these reasons, it may be prudent to avoid dependence on a single antibody. Therefore, both antibodies may be utilised to enumerate or analyse absolute or relative numbers of intermediate and superficial squamous cells in normal or pathological conditions.

Tissue Distribution

15	Kidney, pancreas:	Negative
	liver and endometrium :	
	Colon:	Mab 9G5 - ; Mab HG3 +
	Tonsil:	Stratified epithelium +
20	Epidermis (foreskin):	Superficial and intermediate cells +
	Placenta :	Amnion and cytotrophoblast in placental; membranes + Synciotrophoblast +

25 [In the first trimester, these trophoblast populations were either negative or only weakly stained.]

Biochemical

Mab 9G5 cross-reacts with amnion and placental trophoblast epithelium. On immunoblots, it detects a prominent 40KDa component under both reducing and non-reducing conditions, from detergent extracted material. 5 Therefore, the 9G5 target epitope is likely to be on a non-cytokeratin monomeric protein, which is not conformation dependent. Micro-sequence analysis indicates that the N-terminus is blocked by a methionine residue; further sequence analysis is in 10 progress on proteolytic digests of the intact molecule. Mab HG3 detects a 180 KDa component under reducing and non-reducing conditions, in both detergent soluble and membrane preparations of both amnion and cervix. Therefore, its target epitope is also likely to be a 15 conformationally independent monomeric protein.

EXAMPLE 4 - Mab BC4

This antibody of isotype IgM, was raised against epithelial cells isolated from premalignant cervical 20 biopsies containing CIN.

Reactivity

The antibody primarily reacts with parabasal and intermediate cells in the cervix. 25

(a) Normal cervical epithelium

No of tests: 85; No of patients: 44 (with TZ: 24)

Typically, the reactivity encompasses 2-6 cell

layers above the parabasal layer, and therefore can include the lower layers of intermediate squamous cells. Basal cells, superficial squames and columnar cells are negative.

5 This reactivity pattern, when compared with that of Mabs 6B5, 9G5 and HG3 (see Table 1), enables the intermediate squamous cell population to be enumerated by appropriate deduction.

10 (b) *Premalignant Cervical Epithelium (CIN II/III or III)*

 No of tests: 42; No of patients: 30 (with CIN:
15)

Reactivity on parabasal cells in CIN specimens is
15 either absent or grossly disrupted in 7/15 specimens. Where there was a complete loss of reactivity, the loss occurred abruptly at the junction between normal and
pre-malignant tissue. Where the reactivity was reduced, it was in terms of the number of immuno-stained cell
20 layers.

(c) *Cervical Carcinoma*

 The antibody reacted with 2/7 squamous cell carcinomas tested; of these one was heterogenous in its
25 reactivity.

 The two adenocarcinomas were unreactive.

Tissue Distribution

	Placenta:	All placental tissue, both first trimester and term are negative.
	Kidney:	Negative
	Pancreas:	Negative
5	Colon:	Negative
	Liver:	Negative
	Endometrium:	Negative
	Buccal cavity:	Parabasal cells and stratified epithelium associated with tonsils are positive.
10	Epidermis: (foreskin)	Parabasal cells are positive.

Biochemical

15 The target epitope was probed on immunoblots from detergent-solubilised extracts and membrane preparations of enzyme disaggregated cervical epithelial tissue. The antibody detected a 200 KDa structure under non-
20 reducing conditions only. The target epitope is therefore likely to be present on a conformationally-dependent, non-cytokeratin, protein.

EXAMPLE 5 - Assessment of cervical cells; comparison with PAP test

25

The work described in the preceding examples has resulted in the generation of a panel of antibodies with reactivities against epithelial cell populations

of the human cervix. The reactivities generally overlap, with the exception of Mab 2C7 which reacts specifically with columnar cells. The overlapping specificities of the other four antibodies (6B5, BC4, 5 9G5 and HG3) demonstrate a continuum of reactivities that parallel the differentiation lineage of cervical squamous epithelial cells themselves. It is most probable that the antibody target epitopes are normal differentiation markers which are expressed and lost as 10 the cells differentiate from basal to terminal superficial squames.

The data herein suggests a scenario wherein with the onset of premalignant (CIN) or neoplastic disease conditions, cells can become arrested at a particular 15 stage of differentiation. This may result in an expansion of a particular cell population expressing a detectable cell marker. For example, the anti-parabasal cell reactivity of Mab 6B5 is expanded in specimens with severe CIN II/III.

20 Similarly, a pathological condition may result in an expanded cell population accompanied by the concomitant loss of a characteristic cell marker. For example, the reactivity of Mab BC4, which also reacts against parabasal cells in normal epithelium, is 25 reduced in CIN samples.

The reactivities of Mabs 9G5 and HG3 against intermediate and superficial squames is markedly reduced in premalignant CIN samples. This could be due

to a reduction in the absolute number of intermediate and superficial squames, as a consequence of arrested differentiation at the parabasal stage. Conversely, it could be due to a loss of the relevant cell markers themselves, as a result of the pathological condition.

It should be noted that an actual expansion or reduction in the absolute numbers of a particular cell population is not a pre-requisite for the purposes of the present invention. Any detectable change in antibody binding relative to established parameters of normality is relevant and of use in the present invention. No particular theory or hypothesis limits the nature and scope of the present invention.

A data bank of numerical parameters pertaining to normal smears establishes the "confines of normality", against which test samples may be compared. Any significant variance from the established parameters indicates a need for individual diagnosis by suitably qualified personnel, to assess the clinical status, ie suspect samples are highlighted using the present invention for further examination.

Experimental results

Reactivity of the antibodies 9G5, HG3, 6B5, BC4 and 2C7 was assessed on normal cervical smear samples and CIN-2/CIN-3 cervical smear samples, which were also analysed by the PAP test. The results are shown in Tables 3 and 4.

Normal cervical smear material was collected in parallel with pathological smear samples in a colposcopy clinic. Following the deposition of the primary smear on a glass slide for Papanicolaou staining, the sampling device, either spatula or brush, was placed in 10mls of Hanks buffered saline and agitated. Samples showing obvious contamination with blood were discarded. The resulting cell suspensions were then washed x2 in this buffer and used to generate cytopspins.

Each cytopspin spot contained approximately 10^4 cells.

The MAb reactivities were detected by an indirect immunostaining technique using a Streptavidin-Biotin, alkaline phosphatase detection system. The chromogen (Fuschin; Dakopatts) produced a red stain. Cell nuclei were counterstained with Mayer's haematoxylin.

Scoring of % cells stained was determined by counting dispersed cells; total cells stained red/total no. nuclei. Cell clumps are often present in such preparations and the reactivity of the Mab with these is indicated separately.

The status of the smear sample, normal or CIN, is given as designated by cytological examination (PAP) of the parallel specimen.

Table 3 shows that a pattern of normality for binding of the five monoclonal antibodies can be perceived. In the CIN samples binding of the same

antibodies deviates from this pattern of normality, as shown in Table 4.

It should be noted that in the case of specimens 6 and 7, visually these samples appeared similar to normal smears. On morphological grounds, the majority of cells were obviously superficial squames and dyskaryotic cells were not apparent. In other specimens, for example, 1, 2, 3 and 10, dyskaryotic cells were clearly evident.

Thus, one can establish a pattern of normality for binding of a panel of antibodies to a sample containing cells of the cervix such that deviation from that pattern in binding of those antibodies to a test sample is indicative of some abnormality which warrants further investigation.

All documents mentioned herein are incorporated by reference.

REFERENCES

- Auersperg N and Hauvryl AP, J Nat Cancer Inst 28: pp 605-627, 1962.
- Auersperg N, J Nat Cancer Inst 43: pp 151-173, 1969.
- Harlow E and Lane D (Eds), Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988.
- Holmes CH et al, J Immunology 144: pp 3099-3105, 1990.
- Houlihan et al, J Immunology 149: pp 668-675, 1992.
- Houlihan JM, PhD Thesis, University of Bristol, 1993.
- Kennet R et al (Eds), Monoclonal Antibodies and T-Cell

Hybridomas (Plenum Press, 1980).

Knapp W (Ed), Leukocyte Typing IV, Academic Press,
London, 1989.

Laemmli UK, Nature 227: pp 133-681.

5 Marchalonis JJ, Biochem J. 113: pp 299-305.

Markwell, Analyt. Biochem. 125; pp 427-432.

Schrier M et al, Hybridoma Techniques, Cold Spring
Harbour Laboratory, 1080.

Work and Work (Eds), Laboratory Techniques in

10 Biochemistry and Molecular Biology, Elsevier, 1979.

Table 1.

SUMMARY OF MONOCLONAL ANTIBODY REACTIVITIES AGAINST
NORMAL CERVICAL EPITHELIAL CELLS IN
BIOPSY PROVEN NORMAL CERVICES.

Target Cell Populations	Monoclonal Antibodies				
	2C7	6B5	BC4	9G5	HG3
Basal	-	+/-	-	-	-
Parabasal	-	+	+	+/-	-
Intermediate Squames	-	-	+/-	+	+
Superficial Squames	-	-	-	+	+
Columnar	+	+	-	-	-

Table 2.

REACTIVITY OF MABS 9G3 AND HG3
ON CERVICAL CARCINOMAS

Tumour	Type	2C7	6B5	BC4	9G5	HG3
A	Squamous	-	+++	-	+/-	-
B	Squamous	-	-	-	-	+++
C	Squamous	-	+++	-	+/-	+++
D	Squamous	-	+	-	-	-
E	Squamous	-	+	+/-	+/-	+/-
F	Squamous	-	+/-	-	-	+++
G	Squamous	-	-	+/-	+/-	+/-
H	Adenoc.	+++	+	-	-	+++
I	Adenoc.	-	+	-	-	+

Squamous = Squamous Cell Carcinoma
Adenoc. = Adenocarcinoma

+/- Heterogenous reactivity defined as areas of both reactivity and unreactivity within a specimen.

+ Positive Reactivity.

+++ Extensive intense reactivity.

Table 3.

REACTIVITY OF MONOCLONAL ANTIBODIES WITH
NORMAL CERVICAL SMEARS.

Specimen	(Squamous)		(Parabasal)		(Columnar)
	9G5 ¹	HG3 ¹	6B5 ²	BC4 ²	2C7 ²
1.	75%	88%	-ve	-ve	-ve
2.	68%	86%	-ve	-ve	-ve
3.	65%	70%	-ve	-ve	>50
4.	75%	85%	-ve	-ve	-ve
5.	62%	79%	-ve	ve	-ve
6.	93%	68%	5	1	-ve
7.	83%	74%	2	-ve	>100
8.	72%	85%	7	3	+ve [§]
9.	68%	90%	-ve	-ve	10
10.	85%	90%	-ve	-ve	-ve

mean 74.6 ± 9.79 81.5 ± 8.22

Footnotes

¹200 cells were counted in the case of 9G5 and HG3.

²Numbers given refer to positive cells in the whole field containing approx. 10⁴ cells.

[§]Clumps of stained cells were observed.

Table 4.

REACTIVITY OF MONOCLONAL ANTIBODIES WITH CERVICAL
SMEARS FROM PRE-MALIGNANT SPECIMENS
WITH CIN-2/CIN-3

Patient	Grade	(Squamous)		(Parabasal)		(Columnar)
		9G5 ¹	HG3 ¹	6B5 ²	BC4 ²	2C7 ²
1.	CIN-3	<5%	<2%	-ve	-ve	+ve [§]
2.	CIN-3	NT	<5%	-ve	-ve	+ve [§]
3.	CIN-3	<20%	ND	>50 ^S	-ve	10 [§]
4.	CIN-3	60%	20%	-ve	-ve	1
5.	CIN-3	50%	96%	-ve	-ve	-ve
6.	CIN-3	75%	73%	10	5	-ve
7.	CIN-3	86%	78%	-ve	-ve	10
8.	CIN-2/3	54%	<10%	>20	2 [§]	-ve
9.	CIN-2/3	59%	52%	-ve	-ve	20
10.	CIN-2	15%	15%	19	-ve	4%
11.	CIN-2	38% ^S	20%	-ve	-ve	1

Footnotes

¹200 cells were counted in the case of 9G5 and HG3.

²Numbers given refer to positive cells in the whole field containing approx. 10⁴ cells.

[§] Clumps of stained cells were observed.

CLAIMS:

1. A method of determining abnormality in a tissue sample containing cells of the cervix, the method comprising determining binding of specific binding substances to the sample and comparing the binding with the pattern of binding of said specific binding substances to a normal cervical cell sample.
2. A method according to claim 1 wherein the specific binding substances include one or more polypeptides each comprising an immunoglobulin antigen binding domain.
3. A method according to claim 2 wherein the specific binding substances include one or more polypeptides each comprising the immunoglobulin antigen binding domain obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 0JG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.
4. A method according to claim 1 or claim 2 wherein the specific binding substances include one or more such substances which compete for binding to cervical tissue with one or more antibodies obtainable from a hybridoma selected from those deposited at the European

Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire SP4 0JG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 5 95020717 and 95020719.

5. A hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, 10 Wiltshire SP4 0JG, United Kingdom on 6 February 1995 ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.

6. Use of a hybridoma of claim 4 and/or 15 immunoglobulin antigen binding domain obtainable therefrom in the obtention of one or more specific binding substances of use in assessment of the state or condition of cervical cells in a tissue sample.

20 7. A specific binding substance including an immunoglobulin antigen binding domain obtainable from a hybridoma selected from those deposited at the European Collection of Animal Cell Cultures (ECACC), Centre for Applied Microbiology & Research, Salisbury, Wiltshire 25 SP4 0JG, United Kingdom on 6 February 1995 under the accession numbers ECACC 95020718, 95020716, 95020720, 95020717 and 95020719.

8. A specific binding substance which competes for

binding to cervical tissue with a specific binding substance according to claim 6.

9. Use of a specific binding substance according to claim 6 or claim 7 for assessment of the nature or condition of cells of the cervix in a tissue sample.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100

United States Patent Application
COMBINED DECLARATION AND POWER OF ATTORNEY

Attorney's Docket Number _____

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled:

"Assessment of Cervical Cells"

the specification of which:

[c] was filed as a PCT international application Number PCT/GB97/02108 on 5 August 1997.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below, and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter and having a filing date before that of the application(s) of which priority is claimed:

COUNTRY	APPLICATION NUMBER	DATE OF FILING	PRIORITY CLAIMED
United Kingdom	9616429.8	5 August 1996	Yes



POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

Harold C Hohbach, Reg No 17,757; Aldo J Test, Reg No 18,048; Thomas O Herbert, Reg No 18,612; Donald N MacIntosh, Reg No 20,316; Jerry G Wright, Reg No 20,165; Edward S Wright, Reg No 24,903; David J Brezner, Reg No 24,774, Richard E Backus, Reg No 22,701; James A Sheridan, Reg No 25,435; Robert B Chickering, Reg No 24,286; Richard F Trecartin, Reg No 31,801; C Michael Zimmerman, Reg No 20,451; Walter H Dreger, Reg No 24,190; Steven F Caserza, Reg No 29,780; Laura L Kulhanjian, Reg No 33,257; Julian Caplan, Reg No 14,785; David C Ashby, Reg No 36,432 and Dolly A Vance, Reg No 39,054, provided that if any one of said attorneys ceases being affiliated with the law firm of Flehr Hohbach Test Albritton & Herbert LLP as partner, employee or of counsel, such attorney's appointment as attorney and all powers derived therefrom shall terminate on the date such attorney ceases being so affiliated.

Direct all telephone calls to David C Ashby at (650) 494-8700, and address all correspondence to:

David C Ashby, Esq
FLEHR HOHBACH TEST ALBRITTON & HERBERT LLP
Suite 3400, Four Embarcadero Center
San Francisco, California 94111

FIRST INVENTOR

Full name: Robert James MASON
Residence: New Chapter, 4 Park Road, Cheveley, Newmarket, Suffolk, CB8 9DF, United Kingdom.
Citizenship: Great Britain
Post office address: New Chapter, 4 Park Road, Cheveley, Newmarket, Suffolk, CB8 9DF, United Kingdom.

SECOND INVENTOR

Full name: Edward William PASCOE
Residence: 2 Shortland, Somersham, Huntingdon, PE17 3XJ, United Kingdom (previously of 23 Main Street, Wardy Hill, Ely, Cambridgeshire, CB9 4NX, United Kingdom).
Citizenship: Great Britain
Post office address: 2 Shortland, Somersham, Huntingdon, PE17 3XJ, United Kingdom.

THIRD INVENTOR

Full name: Christopher Harold HOLMES
Residence: _____

Citizenship: Great Britain
Post office address: University of Bristol, Department of Obstetrics & Gynaecology, St. Michael's Hospital, Southwell Street, Bristol, BS2 8EG, United Kingdom.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that wilful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such wilful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of First Inventor _____ Date _____
(Robert James MASON)

Signature of Second Inventor E. W. Pascoe Date 09/02/99
(Edward William PASCOE)

Signature of Third Inventor _____ Date _____
(Christopher Harold HOLMES)

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

Harold C Hohbach, Reg No 17,757; Aldo J Test, Reg No 18,048; Thomas O Herbert, Reg No 18,612; Donald N MacIntosh, Reg No 20,316; Jerry G Wright, Reg No 20,165; Edward S Wright, Reg No 24,903; David J Brezner, Reg No 24,774, Richard E Backus, Reg No 22,701; James A Sheridan, Reg No 25,435; Robert B Chickering, Reg No 24,286; Richard F Trecartin, Reg No 31,801; C Michael Zimmerman, Reg No 20,451; Walter H Dreger, Reg No 24,190; Steven F Caserza, Reg No 29,780; Laura L Kulhanjian, Reg No 33,257; Julian Caplan, Reg No 14,785; David C Ashby, Reg No 36,432 and Dolly A Vance, Reg No 39,054, provided that if any one of said attorneys ceases being affiliated with the law firm of Flehr Hohbach Test Albritton & Herbert LLP as partner, employee or of counsel, such attorney's appointment as attorney and all powers derived therefrom shall terminate on the date such attorney ceases being so affiliated.

Direct all telephone calls to David C Ashby at (650) 494-8700, and address all correspondence to:

David C Ashby, Esq
FLEHR HOHBACH TEST ALBRITTON & HERBERT LLP
Suite 3400, Four Embarcadero Center
San Francisco, California 94111

FIRST INVENTOR

Full name: Robert James MASON
Residence: New Chapter, 4 Park Road, Cheveley, Newmarket, Suffolk, CB8 9DF, United Kingdom.
Citizenship: Great Britain
Post office address: New Chapter, 4 Park Road, Cheveley, Newmarket, Suffolk, CB8 9DF, United Kingdom.

SECOND INVENTOR

Full name: Edward William PASCOE
Residence: 23 Main Street, Wardy Hill, Ely, Cambridgeshire, CB9 4NX, United Kingdom.
Citizenship: Great Britain
Post office address: 23 Main Street, Wardy Hill, Ely, Cambridgeshire, CB9 4NX, United Kingdom.

THIRD INVENTOR

Full name: Christopher Harold HOLMES
Residence: 13 Sunnyside, Stoke Bishop, Bristol, BS9 1BQ, United Kingdom.
Citizenship: Great Britain
Post office address: University of Bristol, Department of Obstetrics & Gynaecology, St. Michael's Hospital, Southwell Street, Bristol, BS2 8EG, United Kingdom.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that wilful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such wilful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of First Inventor _____ Date _____
(Robert James MASON)

Signature of Second Inventor _____ Date _____
(Edward William PASCOE)

Signature of Third Inventor CH Holmes Date 29.01.99
(Christopher Harold HOLMES)

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50